ANTIOXIDANTS & REDOX SIGNALING Volume 16, Number 12, 2012 © Mary Ann Liebert, Inc. DOI: 10.1089/ars.2011.4400

# Insights into Mitochondrial Dysfunction: Aging, Amyloid- $\beta$ , and Tau-A Deleterious Trio

Karen Schmitt, Amandine Grimm, Anna Kazmierczak, Joanna B. Strosznajder, Jürgen Götz, and Anne Eckert

### **Abstract**

Significance: Alzheimer's disease (AD) is an age-related progressive neurodegenerative disorder mainly affecting elderly individuals. The pathology of AD is characterized by amyloid plaques (aggregates of amyloid-β  $[A\beta]$ ) and neurofibrillary tangles (aggregates of tau), but the mechanisms underlying this dysfunction are still partially unclear. Recent Advances: A growing body of evidence supports mitochondrial dysfunction as a prominent and early, chronic oxidative stress-associated event that contributes to synaptic abnormalities and, ultimately, selective neuronal degeneration in AD. Critical Issues: In this review, we discuss on the one hand whether mitochondrial decline observed in brain aging is a determinant event in the onset of AD and on the other hand the close interrelationship of this organelle with  $A\beta$  and tau in the pathogenic process underlying AD. Moreover, we summarize evidence from aging and Alzheimer models showing that the harmful trio "aging,  $A\beta$ , and tau protein" triggers mitochondrial dysfunction through a number of pathways, such as impairment of oxidative phosphorylation (OXPHOS), elevation of reactive oxygen species production, and interaction with mitochondrial proteins, contributing to the development and progression of the disease. Future Directions: The aging process may weaken the mitochondrial OXPHOS system in a more general way over many years providing a basis for the specific and destructive effects of  $A\beta$  and tau. Establishing strategies involving efforts to protect cells at the mitochondrial level by stabilizing or restoring mitochondrial function and energy homeostasis appears to be challenging, but very promising route on the horizon. Antioxid. Redox Signal. 16, 1456–1466.

### Introduction

GING is an inevitable biological process that results in a Aprogressive structural and functional decline, as well as biochemical alterations that altogether lead to reduced ability to adapt to environmental changes. Although aging is almost universally conserved among all organisms, the molecular mechanisms underlying this phenomenon still remain unclear. There are several theories of aging, in which free radical (oxidative stress), DNA, or protein modifications are suggested to play the major causative role (54, 72). A growing body of evidence supports mitochondrial dysfunction as a prominent and early, chronic oxidative stress-associated event that contributes to synaptic abnormalities in aging and, ultimately, increased susceptibility to age-related disorders including Alzheimer's disease (AD) (58). AD is the most common neurodegenerative disorder among elderly individuals. It accounts for up to 80% of all dementia cases and ranks as the fourth leading cause of death among those above 65 years of age. With the increasing average life span of humans, it is highly probable that the number of AD cases will dangerously raise. The pathology of AD characterized by abnormal formation of amyloid plaques (aggregates of amyloid- $\beta$  [A $\beta$ ]) and neurofibrillary tangles (NFT; aggregates of tau) was shown to be accompanied by mitochondrial dysfunction. However, the mechanisms underlying this dysfunction are poorly understood. There remain several open questions: Is age-related oxidative stress accelerating the NFT and A $\beta$  pathologies? Are these lesions causing oxidative stress themselves? Or are there other mechanisms involved? Within the past years, several mouse models have been developed that reproduce the aging process and diverse aspects of AD. These models help in understanding the age-related pathogenic mechanisms that lead to mitochondrial failure in AD, and in particular the interplay of AD-related cellular modifications within this process (17, 18).

<sup>&</sup>lt;sup>1</sup>Neurobiology Laboratory for Brain Aging and Mental Health, Psychiatric University Clinics, University of Basel, Switzerland.

<sup>&</sup>lt;sup>2</sup>Mossakowski Medical Research Center, Polish Academy of Sciences, Department of Cellular Signaling, Warsaw, Poland.

<sup>&</sup>lt;sup>3</sup>Alzheimer's and Parkinson's Disease Laboratory, Brain and Mind Research Institute, University of Sydney, Camperdown, New South Wales, Australia.

### Mitochondrial Aging—the Beginning of the End in AD?

Mitochondria play a pivotal role in cell survival and death by regulating both energy metabolism and apoptotic pathways; they contribute to many cellular functions, including intracellular calcium homeostasis, the alteration of the cellular reduction-oxidation (redox) potential, cell cycle regulation, and synaptic plasticity (47). They are the "powerhouses of cells," providing energy from nutritional sources via ATP generation, which is accomplished through oxidative phosphorylation (OXPHOS) (65). However, when mitochondria fulfill their physiological function, it is as if Pandora's box has been opened, as this vital organelle contains potentially harmful proteins and biochemical reaction centers; mitochondria are the major producers of reactive oxygen species (ROS) at the same time being susceptible targets of ROS toxicity. Unstable ROS are capable of damaging many types of mitochondrial components; this includes oxidative deterioration of mitochondrial DNA (mtDNA), lipids of the mitochondrial membrane, and mitochondrial proteins, and it is thought that this damage that may accumulate over time from ROS generated from aerobic respiration may play a significant role in aging (Fig. 1). Moreover, it was previously demonstrated that nitrosative stress evoked by increased nitric oxide synthesis also leads to protein oxidation as well as mitochondrial and DNA damage, which are common mechanisms occurred in elderly (13, 34, 70).

Although most mitochondrial proteins are encoded by the nuclear genome, the mitochondrial genome encodes proteins required for 13 polypeptide complexes of the respiratory chain involved in ATP synthesis. Given that mtDNA exists in the inner matrix and this is in close proximity to the inner membrane where electrons can form unstable compounds, mtDNA, unlike nuclear DNA (nDNA), is not protected by histones (4) making it more vulnerable to oxidative stress and its mutation rate is about 10-fold higher than that of nDNA, especially in tissues with a high ATP demand like the brain (54). These mtDNA mutations occur in genes encoding electron transport chain (ETC) subunits including NADH dehydrogenase, cytochrome c oxidase (COX), and ATP synthase (83). Eventually, ROS-related mtDNA mutations can result in the synthesis of mutant ETC proteins that, in turn, can lead to the leakage of more electrons and increased ROS production. This so-called "vicious cycle" is hypothesized to play a critical role in the aging process according to the mitochondrial theory of aging. In addition to age-associated increase in mtDNA mutations, the amount of mtDNA also declines with age in various human and rodent tissues (2, 68). Furthermore, abundance of mtDNA correlates with the rate of mitochondrial ATP production (68), suggesting that age-related mitochondrial dysfunction in muscle is related to reduced mtDNA abundance. However, age-associated change in mtDNA abundance seems to be tissue specific, as several studies have reported no change in mtDNA abundance with age in other than muscular tissues in both man and mouse (20, 46).

How does the somatic mtDNA involved in aging phenotypes contribute to AD development? As only a small fraction of AD is caused by autosomal dominant mutations, this comes down to the question of what is causing the prevalent sporadic cases in the first place. Somatic mutations in mtDNA could cause energy deficiency, increased oxidative stress, and accumulation of  $A\beta$ , which act in a vicious cycle reinforcing

mtDNA damage and oxidative stress (45). Indeed, defects in mtDNA associated with decreased cytochrome oxidase activity have been found in AD patients (9). Although a similarly impaired mitochondrial function and subsequent compensatory response have been observed in both non-demented aged and AD subjects, no clear causative mutations in the mtDNA have been correlated to AD; although some variations have functional consequences, including changes in enzymatic activity (40). Perhaps the main differences are that, in AD brains, defects are more profound due to A $\beta$  and tau accumulation, because of decreased compensatory response machinery (Fig. 1).

Many investigators have developed models for studying mitochondrial-related aging (36). Among them senescenceaccelerated mice (SAM) strains are especially useful models to understand the mechanisms of the age-related mitochondrial decline. Behavioral studies showed that learning and memory deficits already started as early as 6 months and worsened with aging in SAMP8 mice (accelerated senescence-prone 8) (53, 77). Moreover, Omata and collaborators showed agerelated changes in cerebral energy production in the 2-monthold SAMP8 followed by a decrease in mitochondrial function compared with SAMR1 mice (accelerated senescence-resistant 1) (51). Aging is not only connected with increased mitochondrial ROS production due to ETC impairment but also with a dysbalance of the protective antioxidant machinery inside mitochondria. For instance, age-related changes in levels of antioxidant enzymes, such as copper/zinc superoxide dismutase (Cu/Zn-SOD) and manganese SOD (Mn-SOD), have been found in liver and cortex of SAMP8 mice when compared with age-matched SAMR1 mice, supporting increased oxidative stress as a key mechanism involved in the aging process (37). More recently, Yew and collaborators have shown an impairment of mitochondrial functions including a decrease of COX activity, mitochondrial ATP content, and mitochondrial glutathione (GSH) level at a relatively early age in SAMP8 mice compared with SAMR1 mice (67, 78). Furthermore, the biochemical consequences of aging have been investigated using proteomic analysis in the brain of SAMP8 and SAMR1 mice at presymptomatic (5-month old) and symptomatic (15-month old) stages (84), revealing differentially expressed proteins with age in both mouse strains, such as Cu/Zn-SOD. Besides the progressive mitochondrial decline and increased oxidative stress, tau hyperphosphorylation was also observed at an early age in the brain of SAMP8 mice (1, 71). In addition, SAMP8 mice showed an agerelated increase in mRNA and protein levels of amyloid-β precursor protein (APP). The cleavage product  $A\beta$  was significantly increased at 9 months in SAMP8 and amyloid plaques started to form at around 16 months of age (48, 73). Altogether, these data indicate that mitochondrial dysfunction is a highly relevant event in the aging process, which is also known as the primary risk factor for AD and other prevalent neurodegenerative disorders.

### Age-Related A $\beta$ and Tau Effects on Mitochondria in AD

AD is a progressive, neurodegenerative disorder, characterized by an age-dependent loss of memory and an impairment of multiple cognitive functions. From a genetic point of view, AD can be classified into two different forms: rare familial forms (FAD) where the disease onset is at an age below

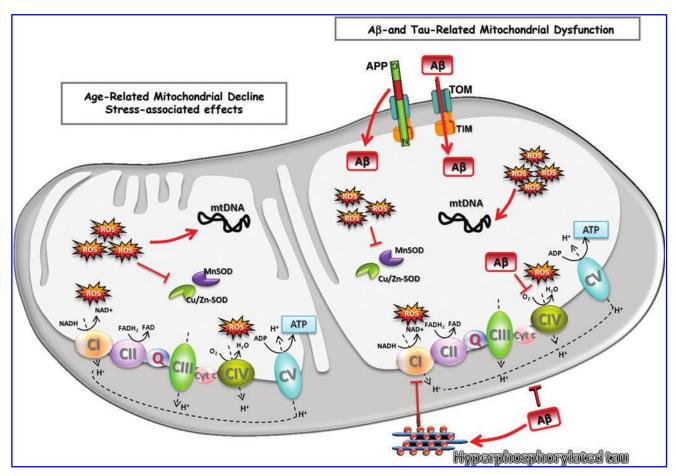


FIG. 1. Aging, Aβ, and tau: toxic consequence on mitochondria. The aging process may weaken the mitochondrial OXPHOS (oxidative phosphorylation) system in a more general way by the accumulation of ROS-induced damage over many years thereby sowing the seeds for specific and destructive effects of  $A\beta$  and tau. ROS induce peroxidation of several mitochondrial macromolecules, such as mtDNA and mitochondrial lipids, contributing to mitochondrial impairment in the mitochondrial matrix. In AD, mitochondria were found to be a target of  $A\beta$  toxicity, which may act directly or indirectly on several proteins, leading to mitochondrial dysfunction. Indeed,  $A\beta$  was found in the OMM and IMM as well as in the matrix. The interaction of  $A\beta$  with the OMM might affect the transport of nuclear-encoded mitochondrial proteins, such as subunits of the ETC CIV, into the organelle via the TOM import machinery. A $\beta$  seems to be able to enter into the mitochondrial matrix through TOM and TIM or could be derived from mitochondria-associated APP metabolism. The interaction of A $\beta$  with the IMM would bring it into contact with respiratory chain complexes with the potential for myriad effects on cellular metabolism. It may be that  $A\beta$  by these interactions affects the activity of several enzymes decreasing the ETC enzyme CIV, reducing the amount of hydrogen that is translocated from the matrix to the intermembrane space, thus impairing the MMP. The dysfunction of the ETC leads to a decreased CV activity and so to a lower ATP synthesis, in addition to an increased ROS production. Interestingly, deregulation of CI is mainly tau dependent, while deregulation of CIV is  $A\beta$  dependent, at both the protein and activity level. A $\beta$ , amyloid- $\beta$ ; AD, Alzheimer's disease; APP, amyloid precursor protein; CI, complex I; CII, complex II; CIII, complex III; CIV, complex IV; CV, complex V, Cu/Zn-SOD, copper/zinc superoxide dismutase; cyt c, cytochrome c; ETC, electron transport chain; IMM, inner mitochondrial membrane; MMP, mitochondrial membrane potential; MnSOD, manganese superoxide dismutase; mtDNA, mitochondrial DNA; OMM, outer mitochondrial membrane; ROS, reactive oxygen species; TIM, translocase of the inner membrane; TOM, translocase of the outer membrane. (To see this illustration in color the reader is referred to the Web version of this article at www.liebertonline.com/ars).

60 years (<1% of the total number of AD case) and the vast majority of sporadic AD cases where onset occurs at an age over 60 years. Genetic studies in FAD patients have identified autosomal dominant mutations in three different genes, encoding the APP (over 20 pathogenic mutations identified) and the presenilins PS1 and PS2 (more than 130 mutations identified) (26). These mutations are directly linked to the increased production of  $A\beta$  from its precursor protein APP, suggesting a direct and pathological role for  $A\beta$  accumulation in the development of AD.

Mitochondrial dysfunction has been proposed as an underlying mechanism in the early stages of AD, since energy deficiency is a fundamental characteristic feature of AD brains (44) as well as of peripheral cells derived from AD patients (22). Understanding the molecular pathways by which the various pathological alterations including  $A\beta$  and tau compromise neuronal integrity, leading to clinical symptoms, has been a long-standing goal of AD research. The successful development of mouse models that mimic diverse aspects of the AD process has facilitated this effort and assisted in

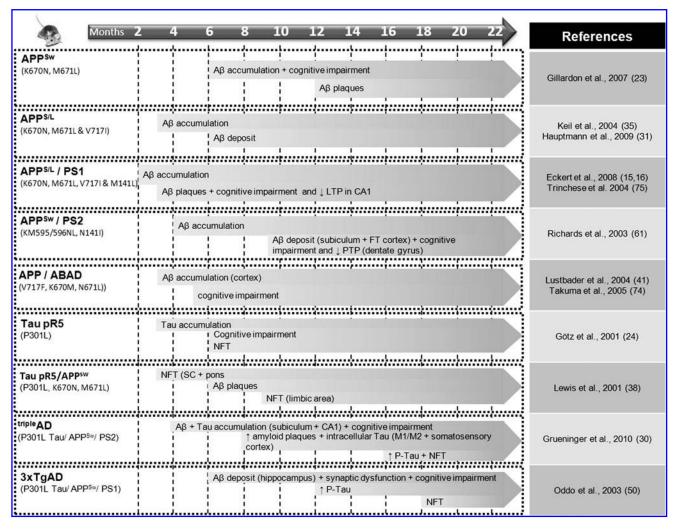


FIG. 2. Age-dependent appearance of histopathological hallmarks in transgenic AD mouse model.

understanding of the age-dependent interplay of A $\beta$  and tau on bioenergetics processes *in vivo* (Figs. 2 and 3).

### Separate modes of AB and tau toxicity on mitochondria

Mitochondria were found to be a target for APP toxicity as both the full-length protein and A $\beta$  accumulate in the mitochondrial import channels, and both lead to mitochondrial dysfunction (7, 42, 55, 56). Several evidences from cellular and animal AD models indicate that  $A\beta$  triggers mitochondrial dysfunction through a number of pathways such as impairment of OXPHOS, elevation of ROS production, interaction with mitochondrial proteins, and alteration of mitochondrial dynamics (52). Indeed, abnormal mitochondrial dynamics have been identified in sporadic and familial AD cases (43, 76) as well as in AD mouse model (6); a distortion probably mediated by altered expression of dynamin-like protein 1 (DLP1), a regulator of mitochondrial fission and distribution, due to elevated oxidative and/or A $\beta$ -induced stress. This modification can disturb the balance between fission and fusion of mitochondria in favor of mitochondrial fission followed by mitochondrial depletion from axons and dendrites and, subsequently, synaptic loss.

Success in developing mouse models that mimic diverse facets of the disease process has greatly facilitated the understanding of physiopathological mechanisms underlying AD. Thus, in 1995, Games and collaborators established the first APP mice model (called PDAPP) bearing the human "Indiana" mutation of the APP gene (V171F). They observed the accumulation of A $\beta$  in the brain and subsequent amyloid plaque formation as well as astrocytosis and neuritic dystrophy (21). Interestingly, in this model cognitive deficits, such as spatial learning impairment, occur before the formation of  $A\beta$ plaques and increase with age (8). This phenomenon was also observed in Tg2576 transgenic mice bearing the human Swedish mutation of the APP gene (K670N, M671L). In fact, in most of the APP mouse models, the cognitive impairment begins concomitantly with A $\beta$  oligomer formation in the brain (around 6 months of age), while neuritic amyloid deposits become visible only between 12 and 23 months and then the amount of deposits increases (23, 31, 35). Thus, memory deficits seem to directly correlate with the accumulation of intracellular A $\beta$  oligomers and not with amyloid plaque formation. Crossing APP transgenic mice with those bearing a mutation in presenilin 1 gene enabled an earlier onset of amyloid plaques compared with APP mice. In one of the most

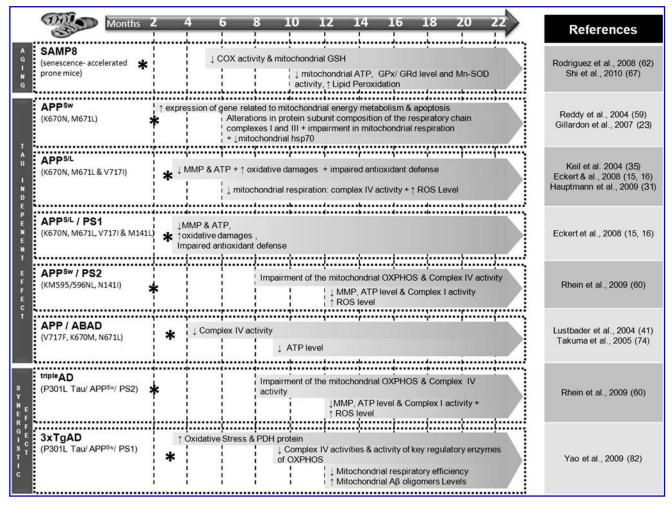


FIG. 3. Age-dependent mitochondrial dysfunction in senescence-accelerated and transgenic AD mouse models. (Star: start of the experiments).

aggressive models, double-transgenic APP<sup>S/L</sup>/PS1 (APP<sup>Swedish/London</sup>/PS1<sup>M141L</sup>) mice, A $\beta$  accumulation begins very soon at 1–2 months of age while cognitive deficits and amyloid plaque formation are already observed at 3 months (3, 16). A stronger decrease of mitochondrial membrane potential as well as ATP level was also found in these mice.

Mitochondrial dysfunctions also appear to a very early stage in these transgenic mouse models. For example, in the APP<sup>Sw</sup> transgenic strain Tg2576, an upregulation of genes related to mitochondrial energy metabolism and apoptosis was observed already at 2 months of age. Alterations in composition of the mitochondrial respiratory chain complexes I and III protein subunit as well as impairment of mitochondrial respiration were detected around 6 months, when soluble  $A\beta$  accumulated in the brain without plaque formation (10, 23, 59). To test the hypothesis that oxidative stress can underlie the deleterious effects of PS mutations, Schuessel and collaborators analyzed lipid peroxidation products (4hydroxynonenal [HNE] and malondialdehyde) and antioxidant defense mechanisms in brain tissue and ROS levels in splenic lymphocytes from transgenic mice bearing the human PS1 M146L mutation (PS1M146L) compared with those from mice transgenic for wild-type human PS1 (PS1wt) and nontransgenic littermate control mice (66). In brain tissue, HNE levels were increased only in aged (19-22 months) PS1M146L transgenic animals compared with PS1wt mice and not in young (3-4 months) or middle-aged mice (13-15 months). Similarly, in splenic lymphocytes expressing the transgenic PS1 proteins, mitochondrial and cytosolic ROS levels were significantly elevated compared with controls only in cells from aged PS1M146L animals. Antioxidant defense mechanisms (activities of antioxidant enzymes including Cu/Zn-SOD, GSH peroxidase, and GSH reductase) as well as susceptibility to oxidative stress in vitro were unaltered. In summary, these results demonstrate that the PS1M146L mutation increases mitochondrial ROS formation and oxidative damage selectively in aged mice. Consistent with this observation, in Swedish amyloid precursor protein (APPSw)/PS2 double-transgenic mice, mitochondrial impairment was first detected at 8 months of age, before amyloid plaque deposition, but after soluble  $A\beta$  accumulation (60, 61). Taken together, these findings are consistent with the recently proposed hypothesis of the age-related A $\beta$ toxicity cascade that suggests that the most toxic  $A\beta$  species that cause majority of molecular and biochemical abnormalities are in fact intracellular soluble oligomeric

aggregates rather than the extracellular, insoluble plaques that may comprise the form of cellular defense against toxicity of oligomers (19). Interestingly, human amylin that aggregates in type 2 diabetic pancreas and shares with  $A\beta$  its amyloidogenic properties also causes an impaired complex IV activity, whereas nonamyloidogenic rat amylin did not (39).

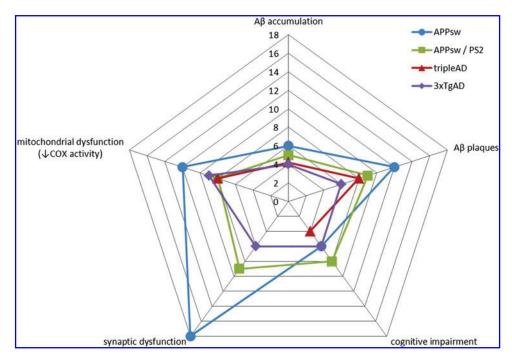
How does tau interfere with mitochondrial function? In its hyperphosphorylated form, tau, which forms the NFTs, the second hallmark lesion in AD, has been shown to block mitochondrial transport, which results in energy deprivation and oxidative stress at the synapse and, hence, neurodegeneration (27, 33, 57). Till now, no mutations in microtubuleassociated protein tau (MAPT) coding genes have been detected in relation to familial forms of AD. However, in familial frontotemporal dementia (FTD) with parkinsonism, mutations in the MAPT gene were identified on chromosome 17. This was the basis for creating a robust mouse model for tau pathology in 2001. These P301L tau-expressing pR5 mice (longest four-repeat 4R2N) show an accumulation of tau as soon as 3 months of age and develop NFTs around 6 months of age (24). A mass spectrometric analysis of the brain proteins from these mice revealed mainly a deregulation of mitochondrial respiratory chain complex components (including complex V), antioxidant enzymes, and synaptic protein space (11). The reduction in mitochondrial complex V levels in the P301L tau mice that was revealed using proteomics was also confirmed as decreased in human P301L FTDP-17 (FTD with parkinsonism linked to chromosome 17) brains. The functional analysis demonstrated age-related mitochondrial dysfunction, together with reduced NADHubiquinone oxidoreductase (complex I) activity as well as age-related impaired mitochondrial respiration and ATP synthesis in pR5 mice model. Mitochondrial dysfunction was also associated with higher levels of ROS in aged transgenic mice. Increased tau pathology resulted in modification of lipid peroxidation levels and the upregulation of antioxidant enzymes in response to oxidative stress (11). Thus, this evidence demonstrated for the first time that not only A $\beta$  but also tau pathology leads to metabolic impairment and oxidative stress by distinct mechanisms from that caused by  $A\beta$  in AD.

### Synergistic modes of Aβ and tau toxicity on mitochondria

Although  $A\beta$  and tau pathologies are both known hallmarks of AD, the mechanisms underlying the interplay between plaques and NFTs (or A $\beta$  and tau, respectively) have remained unresolved. However, a close relationship between mitochondrial impairment and A $\beta$  on the one hand and tau on the other hand has been already established. How do both AD features relate to each other? Is it possible that these two molecules synergistically affect mitochondrial integrity? Several studies suggest that A $\beta$  aggregates and hyperphosphorylated tau may block the mitochondrial carriage to the synapse leading to energy deficiency and neurodegeneration (28). Moreover, the enhanced tau levels may inhibit the transport of APP into axons and dendrites, which suggests a direct link between tau and APP in axonal failure (14, 69). Remarkably, intracerebral A $\beta$  injections amplify a preexisting tau pathology in several transgenic mouse models (5, 25, 29), whereas lack of tau abrogates A $\beta$  toxicity (32, 33). Our findings indicate that in tau transgenic pR5 mice, mitochondria display an enhanced vulnerability toward an A $\beta$  insult *in vitro* (12, 15, 16), suggesting a synergistic action of tau and A $\beta$ pathology on this organelle (Figs. 2 and 3). The A $\beta$  caused a significant reduction of mitochondrial membrane potential in cerebral cells from pR5 mice (11). Furthermore, incubation of isolated mitochondria from pR5 mice with either oligomeric or fibrillar A $\beta$  species resulted in an impairment of the mitochondrial membrane potential and respiration. Interestingly, aging particularly increased the sensitivity of mitochondria to oligomeric A $\beta$  insult compared with that of fibrillar A $\beta$  (15). This suggests that while both oligomeric and fibrillar A $\beta$ species are toxic, they exert different degrees of toxicity. Crossing P301L mutant tau transgenic JNPL3 mice (shortest four-repeat [4R0N] tau together with the P301L mutation) with APP<sup>Sw</sup> transgenic Tg2576 mice revealed the presence of NFT pathology in spinal cord and pons already at 3 months of age (38). A $\beta$  plaques were detected at the age of 6 months and had the same morphology and distribution than in the 1-yearold Tg2576 mice. Taken together, these studies illustrate the existence of a complex interplay between the two key proteins in AD.

Additionally, in recent years triple-transgenic mouse models have been established that combine  $A\beta$  and tau pathologies (Figs. 2 and 3). In these models the contribution of both AD-related proteins on the mitochondrial respiratory machinery and energy homeostasis has been investigated in vivo. Indeed, our group demonstrated a mitochondrial dysfunction in a novel tripletransgenic mouse model (pR5/APPSw/PS2 N141I)—tripleAD mice—using proteomics followed by functional validation (60). Particularly, deregulation of activity of complex I was found to be tau dependent, whereas deregulation of complex IV was A $\beta$  dependent, in 10-month-old <sup>triple</sup>AD mice. The convergent effects of  $A\beta$  and tau led already at the age of 8 months to a depolarization of mitochondrial membrane potential in triple AD mice. Additionally, we found that agerelated oxidative stress also plays a significant part in the deleterious vicious cycle by exaggerating A $\beta$ - and tauinduced disturbances in the respiratory system and ATP synthesis, finally leading to synaptic failure.

Our data complement those obtained in another tripletransgenic mouse model 3xTg-AD (P301Ltau/APP<sup>Sw</sup>/PS1 M146L) (50). In these studies, mitochondrial dysfunction was evidenced by an age-related decrease in the activity of regulatory enzymes of OXPHOS such as COX, or of the Krebs cycle such as pyruvate dehydrogenase, analyzing 3xTg-AD mice aged from 3 to 12 months (82). Besides, these mice also exhibited increased oxidative stress and lipid peroxidation. Most of the effects on mitochondria were seen at the age of 9 months, whereas mitochondrial respiration was significantly decreased at 12 months of age. Importantly, mitochondrial bioenergetics deficits were found to precede the development of AD pathology in the 3xTg-AD mice. Figure 4 nicely shows that AD-specific changes including cognitive impairments,  $A\beta$  accumulation,  $A\beta$  plaques, and mitochondrial dysfunction seem to occur at an earlier onset from single, double up to triple AD transgenic mice models. Together, our studies highlight the key role of mitochondria in AD pathogenesis and consolidate the notion that a synergistic effect of tau and  $A\beta$  enhances the pathological weakening of mitochondria at an early stage of AD.



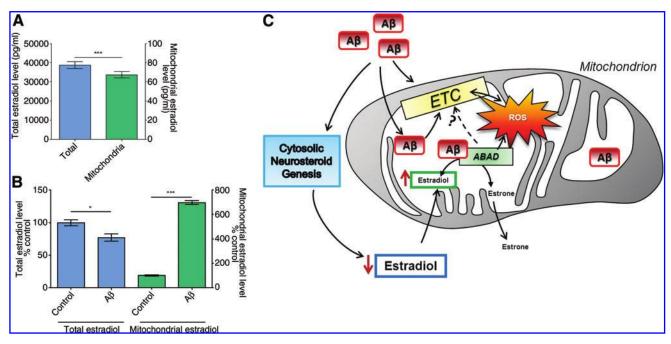
**FIG. 4.** Age-dependent onset of AD-associated pathological changes in different AD mouse models (age in months). In both triple Tg mouse models [Triple AD, (60); 3xTgAD, (50–82)] an earlier onset in the appearance of AD-related changes in the brain can be detected when compared with double transgenic [APP<sup>Sw</sup>/PS2 (60, 61)] and to mice bearing only APP mutations [APP<sup>Sw</sup>, (24)], suggesting again a synergistic effect of Aβ and tau in the pathogenesis of AD. Age of the mice is given in months. APP<sup>Sw</sup>, APP Swedish transgenic mice; APP<sup>Sw</sup>/PS2, APP Swedish/presenilin 2 transgenic mice; tripleAD, APP Swedish/presenilin 2/P301L tau transgenic mice; 3xTgAD, APP Swedish/presenilin 1/P301L tau transgenic mice. (To see this illustration in color the reader is referred to the Web version of this article at www.liebertonline.com/ars).

## $A\beta$ -Binding Alcohol Dehydrogenase: A New Lead to Decode the Mechanisms of $A\beta$ -Induced Mitochondrial Dysfunction

A few years ago, Yan and collaborators showed that the A $\beta$ peptide can directly bind a mitochondrial enzyme called A $\beta$ binding alcohol dehydrogenase (ABAD) that is overexpressed in the brains of Alzheimer's patients and AD mouse models (79). The interaction of A $\beta$  with this enzyme exacerbates mitochondrial dysfunction induced by  $A\beta$  (decrease of mitochondrial complex IV activity, diminution of O2 consumption, and increase of ROS), as shown in double-transgenic mice overexpressing mutant APP and ABAD (81). Furthermore, these mice presented an earlier onset of cognitive impairment and histopathological changes when compared with APP mice, suggesting that the A $\beta$ -ABAD interaction is an important mechanism underlying A $\beta$  toxicity. The A $\beta$ -ABAD complex could have a direct effect on the ETC because ABAD was found to be one of three proteins that comprise the fully functional mammalian mitochondrial RNAse P (63), a function that may not require dehydrogenase activity and that links ABAD directly to the production of mitochondrial ETC proteins and ROS generation.

Recently, it has been shown that inhibition of  $A\beta$ –ABAD interaction by a decoy peptide can restore mitochondrial deficits and improve neuronal and cognitive function (81). Our findings, using SH-SY5Y neuroblastoma cells treated with  $A\beta_{1-42}$ , a cellular model of AD, seem to confirm these observations (Lim *et al.*, unpublished observations). We employed a novel small ABAD-specific inhibitor to investigate

the role of this enzyme in A $\beta$  toxicity. The inhibitor significantly improved metabolic functions impaired by  $A\beta$ , and specifically reduced A $\beta$ -induced oxidative stress and cell death. Furthermore, we have shown previously that the production of estradiol, a well-known neuroprotective neurosteroid and ABAD substrate, is increased after 24h in the presence of a "nontoxic" concentration of  $A\beta$  and is decreased when using a toxic concentration of this peptide (64), suggesting that  $A\beta$  is able to modulate (directly or indirectly) neurosteroid levels. Accordingly, new findings from our group demonstrate that the levels of estradiol in the cytosol and in mitochondria can differently be influenced by  $A\beta$ peptide (500 nM, 5 days of treatment) (Fig. 5A, B). We observed that cytosolic estradiol is reduced in the presence of  $A\beta$ , but at the same time mitochondrial estradiol load was significantly increased. We suggest that this increase is due to an A $\beta$ -induced decrease of ABAD activity, thus limiting the conversion of estradiol in estrone within mitochondria (Fig. 5C). Inhibition of ABAD activity by  $A\beta$  peptide was already demonstrated by Yan and collaborators (80) using  $17\beta$ -estradiol as substrate of the enzyme. One mechanism that could explain this inhibition is the fact that  $A\beta$ -ABAD interaction changes the conformation of the enzyme, avoiding the binding of the cofactor NAD+, and this reduces the metabolic activity of ABAD (41). However, the total amount of estradiol is about 500-fold higher than in the mitochondrial fraction. Even if  $A\beta$  induced an increase in estradiol within mitochondria, the reduction of total estradiol level by other enzymes of the complex steroidogenic pathway may therefore be more relevant for cellular dysfunction. Besides,



**FIG. 5. Aβ–ABAD** interaction and estradiol level in mitochondria. (A) Mitochondrial estradiol is very low compared with total estradiol level in SH-SY5Y neuroblastoma cells. Unpaired t-test, \*\*\*p<0.001. (B) Estradiol level is differently influenced by A $\beta$  peptide in the cytosol and in mitochondria. Paired t-test, \*p<0.05, \*\*\*p<0.0001. (C) A $\beta$  peptide is able to modulate ABAD function by binding directly to this mitochondrial enzyme. This results in the decrease of ABAD-induced conversion of estradiol into estrone with a concomitant increase of ROS levels and an impairment of the ETC in mitochondria. ABAD, A $\beta$ -binding alcohol dehydrogenase. (To see this illustration in color the reader is referred to the Web version of this article at www.liebertonline.com/ars).

it was also speculated that estradiol exhibits a "prooxidant effect" in the presence of ongoing oxidative stress (49). Thereby, estradiol is hydroxylated to catecholestrogens that can enter a redox cycle generating superoxide radicals, leading to a continuous formation of ROS that amplifies oxidative stress.

Thus, inhibition of the A $\beta$ -ABAD interaction seems to be an interesting therapeutic target to block or prevent A $\beta$ -induced mitochondrial toxicity because it could normalize the imbalance between ROS and estradiol levels in mitochondria and thereby help in improving mitochondrial and neuronal function.

### Conclusion

We discuss here the recent findings regarding the possible shared mechanisms involving mitochondrial decline driven by brain aging and the close interrelationship of this organelle with the two main pathological features in the pathogenic process underlying AD.

According to the mitochondrial aging theory, ROS-induced damage and mtDNA mutations accumulate over time inducing ETC impairment and weaken mitochondria function in a rather unspecific way; thus, laying the ground for the two common hallmarks of AD, plaques and NFTs, or A $\beta$  and tau, respectively, which destruct independently as well as synergistically this vital organelle via specific mode of actions on complexes I and IV.

Given the complexities of AD, the key role of mitochondrial dysfunction in the early pathogenic pathways by which  $A\beta$ 

leads to neuronal dysfunction in AD is particularly challenging with respect to establishing therapeutic treatments. Besides the modulation and/or removal of both  $A\beta$  and tau pathology, strategies involving efforts to protect cells at the mitochondrial level by stabilizing or restoring mitochondrial function or by interfering with energy metabolism appear to be promising. Transgenic AD mice, and particularly triple-transgenic models that combine both pathologies in an age-dependent manner (Fig. 4), are valuable tools in monitoring therapeutic interventions at the mitochondrial level. Eventually, this may lead to therapies that prevent the progression of the age-related mitochondrial decline thereby reducing the vulnerability to  $A\beta$  and/or tau at an early stage of the disease.

### **Acknowledgments**

This work has been supported by grants from the Swiss National Science Foundation (#31000\_122572), Sciex\_NMS<sup>CH</sup> and Synapsis Foundation.

### References

- Alvarez-Garcia O, Vega-Naredo I, Sierra V, Caballero B, Tomas-Zapico C, Camins A, Garcia JJ, Pallas M, and Coto-Montes A. Elevated oxidative stress in the brain of senescence-accelerated mice at 5 months of age. *Biogerontology* 7: 43–52, 2006.
- 2. Barazzoni R, Short KR, and Nair KS. Effects of aging on mitochondrial DNA copy number and cytochrome c oxidase gene expression in rat skeletal muscle, liver, and heart. *J Biol Chem* 275: 3343–3347, 2000.

- 3. Blanchard V, Moussaoui S, Czech C, Touchet N, Bonici B, Planche M, Canton T, Jedidi I, Gohin M, Wirths O, Bayer TA, Langui D, Duyckaerts C, Tremp G, and Pradier L. Time sequence of maturation of dystrophic neurites associated with Abeta deposits in APP/PS1 transgenic mice. *Exp Neurol* 184: 247–263, 2003.
- Bohr VA. Repair of oxidative DNA damage in nuclear and mitochondrial DNA, and some changes with aging in mammalian cells. Free Radic Biol Med 32: 804–812, 2002.
- Bolmont T, Clavaguera F, Meyer-Luehmann M, Herzig MC, Radde R, Staufenbiel M, Lewis J, Hutton M, Tolnay M, and Jucker M. Induction of tau pathology by intracerebral infusion of amyloid-beta -containing brain extract and by amyloid-beta deposition in APP x Tau transgenic mice. *Am J Pathol* 171: 2012–2020, 2007.
- Calkins MJ, Manczak M, Mao P, Shirendeb U, and Reddy PH. Impaired mitochondrial biogenesis, defective axonal transport of mitochondria, abnormal mitochondrial dynamics and synaptic degeneration in a mouse model of Alzheimer's disease. Hum Mol Genet 20: 4515–4529, 2011.
- Caspersen C, Wang N, Yao J, Sosunov A, Chen X, Lustbader JW, Xu HW, Stern D, McKhann G, and Yan SD. Mitochondrial Abeta: a potential focal point for neuronal metabolic dysfunction in Alzheimer's disease. FASEB J 19: 2040–2041, 2005.
- 8. Chen G, Chen KS, Knox J, Inglis J, Bernard A, Martin SJ, Justice A, McConlogue L, Games D, Freedman SB, and Morris RG. A learning deficit related to age and beta-amyloid plaques in a mouse model of Alzheimer's disease. *Nature* 408: 975–979, 2000.
- Coskun PE, Beal MF, and Wallace DC. Alzheimer's brains harbor somatic mtDNA control-region mutations that suppress mitochondrial transcription and replication. *Proc Natl* Acad Sci U S A 101: 10726–10731, 2004.
- Crouch PJ, Blake R, Duce JA, Ciccotosto GD, Li QX, Barnham KJ, Curtain CC, Cherny RA, Cappai R, Dyrks T, Masters CL, and Trounce IA. Copper-dependent inhibition of human cytochrome c oxidase by a dimeric conformer of amyloid-beta1-42. *J Neurosci* 25: 672–679, 2005.
- David DC, Hauptmann S, Scherping I, Schuessel K, Keil U, Rizzu P, Ravid R, Drose S, Brandt U, Muller WE, Eckert A, and Gotz J. Proteomic and functional analyses reveal a mitochondrial dysfunction in P301L tau transgenic mice. *J Biol Chem* 280: 23802–23814, 2005.
- David DC, Ittner LM, Gehrig P, Nergenau D, Shepherd C, Halliday G, and Gotz J. Beta-amyloid treatment of two complementary P301L tau-expressing Alzheimer's disease models reveals similar deregulated cellular processes. *Pro*teomics 6: 6566–6577, 2006.
- 13. Domek-Lopacinska KU and Strosznajder JB. Cyclic GMP and nitric oxide synthase in aging and Alzheimer's disease. *Mol Neurobiol* 41: 129–137, 2010.
- Ebneth A, Drewes G, Mandelkow EM, and Mandelkow E. Phosphorylation of MAP2c and MAP4 by MARK kinases leads to the destabilization of microtubules in cells. *Cell Motil* Cytoskeleton 44: 209–224, 1999.
- Eckert A, Hauptmann S, Scherping I, Meinhardt J, Rhein V, Drose S, Brandt U, Fandrich M, Muller WE, and Gotz J. Oligomeric and fibrillar species of beta-amyloid (A beta 42) both impair mitochondrial function in P301L tau transgenic mice. J Mol Med (Berl) 86: 1255–1267, 2008.
- Eckert A, Hauptmann S, Scherping I, Rhein V, Muller-Spahn F, Gotz J, and Muller WE. Soluble beta-amyloid leads to mitochondrial defects in amyloid precursor protein and tau transgenic mice. *Neurodegener Dis* 5: 157–159, 2008.

17. Eckert A, Schmitt K, and Gotz J. Mitochondrial dysfunction—the beginning of the end in Alzheimer's disease? Separate and synergistic modes of tau and amyloid-beta toxicity. *Alzheimers Res Ther* 3: 15, 2011.

- Eckert A, Schulz KL, Rhein V, and Gotz J. Convergence of amyloid-beta and tau pathologies on mitochondria in vivo. Mol Neurobiol 41: 107–114, 2010.
- Fernandez-Vizarra P, Fernandez AP, Castro-Blanco S, Serrano J, Bentura ML, Martinez-Murillo R, Martinez A, and Rodrigo J. Intra- and extracellular Abeta and PHF in clinically evaluated cases of Alzheimer's disease. *Histol Histopathol* 19: 823–844, 2004.
- Frahm T, Mohamed SA, Bruse P, Gemund C, Oehmichen M, and Meissner C. Lack of age-related increase of mitochondrial DNA amount in brain, skeletal muscle and human heart. *Mech Ageing Dev* 126: 1192–1200, 2005.
- Games D, Adams D, Alessandrini R, Barbour R, Berthelette P, Blackwell C, Carr T, Clemens J, Donaldson T, Gillespie F, et al. Alzheimer-type neuropathology in transgenic mice overexpressing V717F beta-amyloid precursor protein. Nature 373: 523–527, 1995.
- Gibson GE, Sheu KF, and Blass JP. Abnormalities of mitochondrial enzymes in Alzheimer disease. J Neural Transm 105: 855–870, 1998.
- Gillardon F, Rist W, Kussmaul L, Vogel J, Berg M, Danzer K, Kraut N, and Hengerer B. Proteomic and functional alterations in brain mitochondria from Tg2576 mice occur before amyloid plaque deposition. *Proteomics* 7: 605–616, 2007.
- Gotz J, Chen F, Barmettler R, and Nitsch RM. Tau filament formation in transgenic mice expressing P301L tau. J Biol Chem 276: 529–534, 2001.
- Gotz J, Chen F, van Dorpe J, and Nitsch RM. Formation of neurofibrillary tangles in P3011 tau transgenic mice induced by Abeta 42 fibrils. *Science* 293: 1491–1495, 2001.
- Gotz J and Ittner LM. Animal models of Alzheimer's disease and frontotemporal dementia. Nat Rev Neurosci 9: 532–544, 2008.
- Gotz J, Ittner LM, Fandrich M, and Schonrock N. Is tau aggregation toxic or protective: a sensible question in the absence of sensitive methods? J Alzheimers Dis 14: 423–429, 2008.
- Gotz J, Ittner LM, and Kins S. Do axonal defects in tau and amyloid precursor protein transgenic animals model axonopathy in Alzheimer's disease? *J Neurochem* 98: 993–1006, 2006.
- Gotz J, Schild A, Hoerndli F, and Pennanen L. Amyloidinduced neurofibrillary tangle formation in Alzheimer's disease: insight from transgenic mouse and tissue-culture models. *Int J Dev Neurosci* 22: 453–465, 2004.
- 30. Grueninger F, Bohrmann B, Czech C, Ballard TM, Frey JR, Weidensteiner C, von Kienlin M, and Ozmen L. Phosphorylation of Tau at S422 is enhanced by Abeta in TauPS2APP triple transgenic mice. *Neurobiol Dis* 37: 294–306, 2010.
- Hauptmann S, Scherping I, Drose S, Brandt U, Schulz KL, Jendrach M, Leuner K, Eckert A, and Muller WE. Mitochondrial dysfunction: an early event in Alzheimer pathology accumulates with age in AD transgenic mice. Neurobiol Aging 30: 1574–1586, 2009.
- 32. Ittner LM and Gotz J. Amyloid-beta and tau a toxic pas de deux in Alzheimer's disease. *Nat Rev Neurosci* 12: 65–72, 2011.
- 33. Ittner LM, Ke YD, Delerue F, Bi M, Gladbach A, van Eersel J, Wolfing H, Chieng BC, Christie MJ, Napier IA, Eckert A, Staufenbiel M, Hardeman E, and Gotz J. Dendritic function of tau mediates amyloid-beta toxicity in Alzheimer's disease mouse models. *Cell* 142: 387–397, 2010.

- Jesko H, Chalimoniuk M, and Strosznajder JB. Activation of constitutive nitric oxide synthase(s) and absence of inducible isoform in aged rat brain. *Neurochem Int* 42: 315–322, 2003.
- Keil U, Bonert A, Marques CA, Scherping I, Weyermann J, Strosznajder JB, Muller-Spahn F, Haass C, Czech C, Pradier L, Muller WE, and Eckert A. Amyloid beta-induced changes in nitric oxide production and mitochondrial activity lead to apoptosis. J Biol Chem 279: 50310–50320, 2004.
- 36. Kuro-o M. Disease model: human aging. *Trends Mol Med* 7: 179–181, 2001.
- 37. Kurokawa T, Asada S, Nishitani S, and Hazeki O. Agerelated changes in manganese superoxide dismutase activity in the cerebral cortex of senescence-accelerated prone and resistant mouse. *Neurosci Lett* 298: 135–138, 2001.
- 38. Lewis J, McGowan E, Rockwood J, Melrose H, Nacharaju P, Van Slegtenhorst M, Gwinn-Hardy K, Paul Murphy M, Baker M, Yu X, Duff K, Hardy J, Corral A, Lin WL, Yen SH, Dickson DW, Davies P, and Hutton M. Neurofibrillary tangles, amyotrophy and progressive motor disturbance in mice expressing mutant (P301L) tau protein. *Nat Genet* 25: 402–405, 2000.
- 39. Lim YA, Rhein V, Baysang G, Meier F, Poljak A, Raftery MJ, Guilhaus M, Ittner LM, Eckert A, and Gotz J. Abeta and human amylin share a common toxicity pathway via mitochondrial dysfunction. *Proteomics* 10: 1621–1633, 2010.
- Lu J, Wang K, Rodova M, Esteves R, Berry D, Lezi E, Crafter A, Barrett M, Cardoso SM, Onyango I, Parker WD, Fontes J, Burns JM, and Swerdlow RH. Polymorphic variation in cytochrome oxidase subunit genes. *J Alzheimers Dis* 21: 141– 154, 2010.
- 41. Lustbader JW, Cirilli M, Lin C, Xu HW, Takuma K, Wang N, Caspersen C, Chen X, Pollak S, Chaney M, Trinchese F, Liu S, Gunn-Moore F, Lue LF, Walker DG, Kuppusamy P, Zewier ZL, Arancio O, Stern D, Yan SS, and Wu H. ABAD directly links Abeta to mitochondrial toxicity in Alzheimer's disease. *Science* 304: 448–452, 2004.
- 42. Manczak M, Anekonda TS, Henson E, Park BS, Quinn J, and Reddy PH. Mitochondria are a direct site of A beta accumulation in Alzheimer's disease neurons: implications for free radical generation and oxidative damage in disease progression. Hum Mol Genet 15: 1437–1449, 2006.
- 43. Manczak M, Calkins MJ, and Reddy PH. Impaired mitochondrial dynamics and abnormal interaction of amyloid beta with mitochondrial protein Drp1 in neurons from patients with Alzheimer's disease: implications for neuronal damage. Hum Mol Genet 20: 2495–2509, 2011.
- 44. Manczak M, Park BS, Jung Y, and Reddy PH. Differential expression of oxidative phosphorylation genes in patients with Alzheimer's disease: implications for early mitochondrial dysfunction and oxidative damage. *Neuromolecular Med* 5: 147–162, 2004.
- 45. Mao P and Reddy PH. Aging and amyloid beta-induced oxidative DNA damage and mitochondrial dysfunction in Alzheimer's disease: Implications for early intervention and therapeutics. *Biochim Biophys Acta* 1812: 1359–1370, 2011.
- Masuyama M, Iida R, Takatsuka H, Yasuda T, and Matsuki T. Quantitative change in mitochondrial DNA content in various mouse tissues during aging. *Biochim Biophys Acta* 1723: 302–308, 2005.
- Mattson MP, Gleichmann M, and Cheng A. Mitochondria in neuroplasticity and neurological disorders. *Neuron* 60: 748– 766, 2008.
- 48. Morley JE, Kumar VB, Bernardo AE, Farr SA, Uezu K, Tumosa N, and Flood JF. Beta-amyloid precursor polypeptide

- in SAMP8 mice affects learning and memory. *Peptides* 21: 1761–1767, 2000.
- 49. Nilsen J. Estradiol and neurodegenerative oxidative stress. *Front Neuroendocrinol* 29: 463–475, 2008.
- Oddo S, Caccamo A, Shepherd JD, Murphy MP, Golde TE, Kayed R, Metherate R, Mattson MP, Akbari Y, and LaFerla FM. Triple-transgenic model of Alzheimer's disease with plaques and tangles: intracellular Abeta and synaptic dysfunction. *Neuron* 39: 409–421, 2003.
- 51. Omata N, Murata T, Fujibayashi Y, Waki A, Sadato N, Yoshimoto M, Wada Y, and Yonekura Y. Age-related changes in energy production in fresh senescence-accelerated mouse brain slices as revealed by positron autoradiography. *Dement Geriatr Cogn Disord* 12: 78–84, 2001.
- 52. Pagani L and Eckert A. Amyloid-Beta interaction with mitochondria. *Int J Alzheimers Dis* 2011: 925050, 2011.
- 53. Pallas M, Camins A, Smith MA, Perry G, Lee HG, and Casadesus G. From aging to Alzheimer's disease: unveiling "the switch" with the senescence-accelerated mouse model (SAMP8). *J Alzheimers Dis* 15: 615–624, 2008.
- 54. Park CB and Larsson NG. Mitochondrial DNA mutations in disease and aging. *J Cell Biol* 193: 809–818, 2011.
- 55. Pavlov PF, Hansson Petersen C, Glaser E, and Ankarcrona M. Mitochondrial accumulation of APP and Abeta: significance for Alzheimer disease pathogenesis. *J Cell Mol Med* 13: 4137–4145, 2009.
- Pavlov PF, Wiehager B, Sakai J, Frykman S, Behbahani H, Winblad B, and Ankarcrona M. Mitochondrial gammasecretase participates in the metabolism of mitochondriaassociated amyloid precursor protein. FASEB J 25: 78–88, 2011.
- Reddy PH. Abnormal tau, mitochondrial dysfunction, impaired axonal transport of mitochondria, and synaptic deprivation in Alzheimer's disease. *Brain Res* 1415: 136–148, 2011.
- Reddy PH and Beal MF. Amyloid beta, mitochondrial dysfunction and synaptic damage: implications for cognitive decline in aging and Alzheimer's disease. Trends Mol Med 14: 45–53, 2008.
- 59. Reddy PH, McWeeney S, Park BS, Manczak M, Gutala RV, Partovi D, Jung Y, Yau V, Searles R, Mori M, and Quinn J. Gene expression profiles of transcripts in amyloid precursor protein transgenic mice: up-regulation of mitochondrial metabolism and apoptotic genes is an early cellular change in Alzheimer's disease. Hum Mol Genet 13: 1225–1240, 2004.
- 60. Rhein V, Song X, Wiesner A, Ittner LM, Baysang G, Meier F, Ozmen L, Bluethmann H, Drose S, Brandt U, Savaskan E, Czech C, Gotz J, and Eckert A. Amyloid-beta and tau synergistically impair the oxidative phosphorylation system in triple transgenic Alzheimer's disease mice. *Proc Natl Acad Sci U S A* 106: 20057–20062, 2009.
- 61. Richards JG, Higgins GA, Ouagazzal AM, Ozmen L, Kew JN, Bohrmann B, Malherbe P, Brockhaus M, Loetscher H, Czech C, Huber G, Bluethmann H, Jacobsen H, and Kemp JA. PS2APP transgenic mice, coexpressing hPS2mut and hAPPswe, show age-related cognitive deficits associated with discrete brain amyloid deposition and inflammation. *J Neurosci* 23: 8989–9003, 2003.
- 62. Rodriguez MI, Escames G, Lopez LC, Lopez A, Garcia JA, Ortiz F, Sanchez V, Romeu M, and Acuna-Castroviejo D. Improved mitochondrial function and increased life span after chronic melatonin treatment in senescent prone mice. *Exp Gerontol* 43: 749–756, 2008.
- 63. Sapra AK, Anko ML, Grishina I, Lorenz M, Pabis M, Poser I, Rollins J, Weiland EM, and Neugebauer KM. SR protein family members display diverse activities in the formation of nascent and mature mRNPs in vivo. Mol Cell 34: 179–190, 2009.

- 64. Schaeffer V, Meyer L, Patte-Mensah C, Eckert A, and Mensah-Nyagan AG. Dose-dependent and sequence-sensitive effects of amyloid-beta peptide on neurosteroidogenesis in human neuroblastoma cells. *Neurochem Int* 52: 948–955, 2008.
- 65. Scheffler IE. A century of mitochondrial research: achievements and perspectives. *Mitochondrion* 1: 3–31, 2001.
- Schuessel K, Frey C, Jourdan C, Keil U, Weber CC, Muller-Spahn F, Muller WE, and Eckert A. Aging sensitizes toward ROS formation and lipid peroxidation in PS1M146L transgenic mice. Free Radic Biol Med 40: 850–862, 2006.
- 67. Shi C, Xiao S, Liu J, Guo K, Wu F, Yew DT, and Xu J. Ginkgo biloba extract EGb761 protects against aging-associated mitochondrial dysfunction in platelets and hippocampi of SAMP8 mice. *Platelets* 21: 373–379, 2010.
- 68. Short KR, Bigelow ML, Kahl J, Singh R, Coenen-Schimke J, Raghavakaimal S, and Nair KS. Decline in skeletal muscle mitochondrial function with aging in humans. *Proc Natl Acad Sci U S A* 102: 5618–5623, 2005.
- 69. Stamer K, Vogel R, Thies E, Mandelkow E, and Mandelkow EM. Tau blocks traffic of organelles, neurofilaments, and APP vesicles in neurons and enhances oxidative stress. *J Cell Biol* 156: 1051–1063, 2002.
- 70. Strosznajder JB, Jesko H, Zambrzycka A, Eckert A, and Chalimoniuk M. Age-related alteration of activity and gene expression of endothelial nitric oxide synthase in different parts of the brain in rats. *Neurosci Lett* 370: 175–179, 2004.
- 71. Sureda FX, Gutierrez-Cuesta J, Romeu M, Mulero M, Canudas AM, Camins A, Mallol J, and Pallas M. Changes in oxidative stress parameters and neurodegeneration markers in the brain of the senescence-accelerated mice SAMP-8. *Exp Gerontol* 41: 360–367, 2006.
- 72. Swerdlow RH. Brain aging, Alzheimer's disease, and mitochondria. *Biochim Biophys Acta* 1812: 1630–1639, 2011.
- 73. Takemura M, Nakamura S, Akiguchi I, Ueno M, Oka N, Ishi-kawa S, Shimada A, Kimura J, and Takeda T. Beta/A4 proteinlike immunoreactive granular structures in the brain of senescence-accelerated mouse. *Am J Pathol* 142: 1887–1897, 1993.
- 74. Takuma K, Yao J, Huang J, Xu H, Chen X, Luddy J, Trillat AC, Stern DM, Arancio O, and Yan SS. ABAD enhances Abeta-induced cell stress via mitochondrial dysfunction. *FASEB J* 19: 597–598, 2005.
- 75. Trinchese F, Liu S, Battaglia F, Walter S, Mathews PM, and Arancio O. Progressive age-related development of Alzheimer-like pathology in APP/PS1 mice. *Ann Neurol* 55: 801–814, 2004.
- 76. Wang X, Su B, Fujioka H, and Zhu X. Dynamin-like protein 1 reduction underlies mitochondrial morphology and distribution abnormalities in fibroblasts from sporadic Alzheimer's disease patients. *Am J Pathol* 173: 470–482, 2008.
- 77. Watanabe K, Tonosaki K, Kawase T, Karasawa N, Nagatsu I, Fujita M, and Onozuka M. Evidence for involvement of dysfunctional teeth in the senile process in the hippocampus of SAMP8 mice. *Exp Gerontol* 36: 283–295, 2001.
- Xu J, Shi C, Li Q, Wu J, Forster EL, and Yew DT. Mitochondrial dysfunction in platelets and hippocampi of senescence-accelerated mice. *J Bioenerg Biomembr* 39: 195–202, 2007.
- 79. Yan SD, Fu J, Soto C, Chen X, Zhu H, Al-Mohanna F, Collison K, Zhu A, Stern E, Saido T, Tohyama M, Ogawa S, Roher A, and Stern D. An intracellular protein that binds amyloid-beta peptide and mediates neurotoxicity in Alzheimer's disease. *Nature* 389: 689–695, 1997.
- 80. Yan SD, Shi Y, Zhu A, Fu J, Zhu H, Zhu Y, Gibson L, Stern E, Collison K, Al-Mohanna F, Ogawa S, Roher A, Clarke SG, and Stern DM. Role of ERAB/L-3-hydroxyacyl-coenzyme A

- dehydrogenase type II activity in Abeta-induced cytotoxicity. *J Biol Chem* 274: 2145–2156, 1999.
- 81. Yao J, Du H, Yan S, Fang F, Wang C, Lue LF, Guo L, Chen D, Stern DM, Gunn Moore FJ, Xi Chen J, Arancio O, and Yan SS. Inhibition of amyloid-beta (Abeta) peptide-binding alcohol dehydrogenase-Abeta interaction reduces Abeta accumulation and improves mitochondrial function in a mouse model of Alzheimer's disease. *J Neurosci* 31: 2313–2320, 2011.
- 82. Yao J, Irwin RW, Zhao L, Nilsen J, Hamilton RT, and Brinton RD. Mitochondrial bioenergetic deficit precedes Alzheimer's pathology in female mouse model of Alzheimer's disease. *Proc Natl Acad Sci U S A* 106: 14670–14675, 2009.
- 83. Yen TC, Su JH, King KL, and Wei YH. Ageing-associated 5 kb deletion in human liver mitochondrial DNA. *Biochem Biophys Res Commun* 178: 124–131, 1991.
- Zhu L, Yu J, Shi Q, Lu W, Liu B, Xu S, Wang L, Han J, and Wang X. Strain- and age-related alteration of proteins in the brain of SAMP8 and SAMR1 mice. J Alzheimers Dis 23: 641–654, 2011.

Address correspondence to:
Dr. Anne Eckert
Neurobiology Laboratory for Brain Aging and Mental Health
Psychiatric University Clinics
University of Basel
CH-4012 Basel
Switzerland

E-mail: anne.eckert@upkbs.ch

Date of first submission to ARS Central, November 10, 2011; date of acceptance, November 27, 2011.

### **Abbreviations Used**

 $A\beta = \text{amyloid-}\beta$ 

 $ABAD = A\beta$ -binding alcohol dehydrogenase

AD = Alzheimer's disease

APP = amyloid precursor protein

APP<sup>Sw</sup> = Swedish amyloid precursor protein

APPwt = wild-type amyloid precursor protein

COX = cytochrome c oxidase

DLP1 = dynamin-like protein 1

ETC = electron transport chain

FTD = frontotemporal dementia

GSH = glutathione

HA = human amylin

HNE = 4-hydroxynonenal

IMM = inner mitochondrial membrane

MAPT = microtubule-associated protein tau

MMP = mitochondrial membrane potential

mtDNA = mitochondrial DNA

nDNA = nuclear DNA

NFT = neurofibrillary tangle

NO = nitric oxide

OMM = outer mitochondrial membrane

OXPHOS = oxidative phosphorylation

PDH = pyruvate dehydrogenase

PS = presenilin

ROS = reactive oxygen species

SAM = senescence-accelerated mice

SOD = superoxide dismutase

TIM = translocase of the inner membrane

TOM = translocase of the outer membrane

### This article has been cited by:

- 1. Amandine Grimm, Yun-An Lim, Ayikoe Guy Mensah-Nyagan, Jürgen Götz, Anne Eckert. 2012. Alzheimer's Disease, Oestrogen and Mitochondria: an Ambiguous Relationship. *Molecular Neurobiology* **46**:1, 151-160. [CrossRef]
- 2. Joanna B. Strosznajder, Grzegorz A. Czapski, Agata Adamczyk, Robert P. Strosznajder. 2012. Poly(ADP-ribose) Polymerase-1 in Amyloid Beta Toxicity and Alzheimer's Disease. *Molecular Neurobiology* **46**:1, 78-84. [CrossRef]